### **Helix Aortopathies Panel**



Item	Description		
Test Name	Helix Aortopathies Panel		
Test Type	Cardio		
Catalog Number	AORT1		
Procedure Code	H01024-2 (Helix)		
Test Description	This panel evaluates 29 genes associated with inherited aortopathy and related conditions. Genes included are associated with isolated thoracic aortic aneurysms and dissections (TAAD), Marfan Syndrome, and conditions which may include aortopathy as one feature.		
Genes Tested	ACTA2, ADAMTS10, BGN, CBS, COL3A1, COL5A1, COL5A2, EFEMP2, FBN1, FBN2, FLNA, FOXE3, LOX, MED12, MFAP5, MYH11, MYLK, NOTCH1, PLOD1, PRKG1, SKI, SLC2A10, SMAD2, SMAD3, SMAD4, TGFB2, TGFB3, TGFBR1, TGFBR2		
Genetics Information	This test utilizes next-generation sequencing to detect single nucleotide variants, insertions and deletions up to 20 bp, and copy number variants in genes associated with inherited aortopathy and related conditions. Genes included are associated with isolated thoracic aortic aneurysms and dissections (TAAD), Marfan Syndrome, and conditions which may include aortopathy as one feature.		
Indications For Testing	A relevant personal and/or family history suggestive of an inherited aortopathy.		
Clinical Descriptions	Aortopathies refer to a group of genetic disorders that affect the structure and function of the aorta. The genes on this panel include those associated with isolated thoracic aortic aneurysms and dissections (TAAD) as well as syndromic genetic conditions that may include aortopathy as part of the spectrum of disease, such as Marfan syndrome, Loeys-Dietz syndrome or Ehlers-Danlos syndrome.		
Conditions	Arterial tortuosity syndrome (SLC2A10) Contractural arachnodactyly, congenital (FBN2) Ehlers-Danlos syndrome types I, III, IV, VI (COL3A1, COL5A1, COL5A2, PLOD1) Homocystinuria (CBS) Loeys-Dietz syndrome (LDS) (SMAD2, SMAD3, TGFB2, TGFB3, TGFBR1, TGFBR2) Marfan syndrome and other FBN1-related conditions (FBN1) Periventricular nodular heterotopia and other FLNA-related conditions (FLNA) Thoracic aortic aneurysms and aortic dissections (TAAD) (ACTA2, BGN, EFEMP2, FBN1, FOXE3, LOX, MFAP5, MYH11, MYLK, PRKG1)		
Interpretation	All detected variants are evaluated according to American College of Medical Genetics and Genomics recommendations. Variants are classified based on known, predicted, or possible pathogenicity and reported with interpretive comments detailing their potential or known significance.		
Reclassification of Variants	Helix does not systematically review their variant database looking for classification changes. Helix will review the classification of previously reported variants upon request of the ordering physician/provider. Ordering physicians/providers may contact Helix Customer Support or their Dedicated Advisor and request a review of the variant classification to be performed. At the discretion of the laboratory director, the frequency of reclassification requests may be limited to once per year, no earlier than 12 months after initial variant interpretation has been performed.		

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Variant Evaluation	Variant classification is performed using the guidelines set forth by the American College of Medical Genetics and Genomics and the Association for Molecular Pathology, with modifications as suggested by domain specific Expert Panels of the Clinical genome Resource (ClinGen) when available. Variant pathogenicity is categorized as benign, likely benign, variant of uncertain significance (VUS), likely pathogenic, or pathogenic.	
Turnaround Time	7 to 24 days	
Available in NY State	No	
Test Classification	This test was developed, and its performance characteristics determined, by Helix, Inc. in a manner consistent with CLIA requirements. This test has not been cleared or approved by the US Food and Drug Administration.	
Performing Laboratory Information		
Regulatory Information	CLIA Complexity: High Test Classification: Non-Waived/ Laboratory Developed Test	
CLIA Category	Chemistry / Routine Chemistry	

## Methods & Limitations for Helix Aortopathies Panel



Extracted DNA is enriched for targeted regions and then sequenced using the Helix Exome+ (R) assay on an Illumina DNA sequencing system. Data is then aligned to a modified version of GRCh38 and all genes are analyzed using the MANE transcript and MANE Plus Clinical transcript, when available. Small variant calling is completed using a customized version of Sentieon's DNAseq software, augmented by a proprietary small variant caller for difficult variants. Copy number variants (CNVs) are then called using a proprietary bioinformatics pipeline based on depth analysis with a comparison to similarly sequenced samples. Interpretation is based upon guidelines published by the American College of Medical Genetics and Genomics (ACMG) and the Association for Molecular Pathology (AMP) or their modification by ClinGen Variant Curation Expert Panels when available. Interpretation is limited to the transcripts indicated on the report, +/- 10 bp into intronic regions, except as noted below. Helix variant classifications include pathogenic, likely pathogenic, variant of uncertain significance (VUS), likely benign, and benign. Variants classified as pathogenic, likely pathogenic, or VUS are included in the report. All reported variants (except for VUSs with limited evidence of pathogenicity) are confirmed through secondary manual inspection of DNA sequence data or orthogonal testing. Benign and likely benign variants are not reported but are available upon request. Risk estimations and management guidelines included in this report are based on analysis of primary literature and recommendations of applicable professional societies, and should be regarded as approximations.

Based on validation studies, this assay delivers > 99% sensitivity and specificity for single nucleotide variants and insertions and deletions (indels) up to 20 bp. Larger indels and complex variants are also reported but sensitivity may be reduced. Based on validation studies, this assay delivers > 99% sensitivity to multi-exon CNVs and > 90% sensitivity to single-exon CNVs. This test may not detect variants in challenging regions (such as short tandem repeats, homopolymer runs, and segment duplications), sub-exonic CNVs, chromosomal aneuploidy, or variants in the presence of mosaicism. Phasing will be attempted and reported, when possible. Structural rearrangements such as inversions, translocations, and gene conversions are not tested in this assay unless explicitly indicated. Additionally, deep intronic, promoter, and enhancer regions may not be covered. It is important to note that this assay cannot detect all variants known to increase disease risk, and that a negative result does not guarantee that the tested individual does not carry a rare, undetectable variant in genes analyzed. Any potential incidental findings outside of these genes and conditions will not be identified, nor reported. The results of a genetic test may be influenced by various factors, including bone marrow transplantation, blood transfusions, or in rare cases, hematolymphoid neoplasms.

#### **Gene Specific Notes:**

FOXE3: Analysis begins at chr1:47416567 (GRCh38) and excludes the first quarter of exon 1. TGFBR1: Analysis for exon 1 will not be performed.

#### Disclaimer:

This test was developed and validated by Helix, Inc. This test has not been cleared or approved by the United States Food and Drug Administration (FDA). The Helix laboratory is accredited by the College of American Pathologists (CAP) and certified under the Clinical Laboratory Improvement Amendments (CLIA #: 05D2117342) to perform high-complexity clinical tests. This test is used for clinical purposes. It should not be regarded as investigational or for research.

## Targeted Genes & Methodology for Helix Aortopathies Panel



The following applies to the Helix Aortopathies Panel. Testing is performed to evaluate for the presence of variants in coding regions and extending to +/- 10 base pairs of adjacent intronic sequences on either side of the coding exons of the genes analyzed. In addition, the analysis will cover select non-coding variants, as listed below. Next-generation sequencing is performed to test for the presence of small variants and copy number variants in the genes analyzed. Confirmation of select reportable variants may be performed by alternate methodologies based on internal laboratory criteria.

This list is current from November 2024 to the present. This document is intended to highlight additional evaluations for variants of high clinical interest as well as technical limitations. For questions regarding genes, reference transcripts, or specific regions covered, contact Helix Customer Service at (844) 211-2070.

Genomic Build: GRCh38 Catalog Number: AORT1

Gene	Transcript	Additional Evaluations	Technical Limitations
ACTA2	NM_001613.4	_	-
ADAMTS10	NM_030957.4	_	-
BGN	NM_001711.6	_	-
CBS	NM_000071.3	_	_
COL3A1	NM_000090.4	_	-
COL5A1	NM_000093.5	_	_
COL5A2	NM_000393.5	_	_
EFEMP2	NM_016938.5	_	-
FBN1	NM_000138.5	_	_
FBN2	NM_001999.4	_	_
FLNA	NM_001110556.2	_	_
FOXE3	NM_012186.3	_	Analysis begins at chr1:47416567 (GRCh38) and excludes the first quarter of exon 1
LOX	NM_002317.7	_	_
MED12	NM_005120.3	-	_
MFAP5	NM_003480.4	_	_
MYH11	NM_002474.3; NM_001040113.2	_	_

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Gene	Transcript	Additional Evaluations	Technical Limitations
MYLK	NM_053025.4	_	_
NOTCH1	NM_017617.5	_	_
PLOD1	NM_000302.4	_	_
PRKG1	NM_006258.4	_	_
SKI	NM_003036.4	_	_
SLC2A10	NM_030777.4	-	_
SMAD2	NM_005901.6	-	_
SMAD3	NM_005902.4	_	_
SMAD4	NM_005359.6	_	_
TGFB2	NM_003238.6	_	_
TGFB3	NM_003239.5	_	_
TGFBR1	NM_004612.4	_	Analysis for exon 1 will not be performed
TGFBR2	NM_003242.6	_	_